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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<p>(51) International Patent Classification 6 : C07H 21/04, C12P 21/06, 21/02, C12N 1/20, 15/00, C07K 1/00, 14/52</p>	<p>A1</p>	<p>(11) International Publication Number: WO 97/15586 (43) International Publication Date: AC 1 May 1997 (01.05.97)</p>
<p>(21) International Application Number: PCT/US96/16778 (22) International Filing Date: 17 October 1996 (17.10.96) (30) Priority Data: 08/553,727 23 October 1995 (23.10.95) US (71) Applicant: TULARIK, INC. [US/US]; Two Corporate Drive, South San Francisco, CA 94080 (US). (72) Inventors: BAICHWAL, Vijay, R.; Two Corporate Drive, South San Francisco, CA 94080 (US). HUANG, Jianing; Two Corporate Drive, South San Francisco, CA 94080 (US). HSU, Hailing; Two Corporate Drive, South San Francisco, CA 94080 (US). GOEDEL, David, V.; Two Corporate Drive, South San Francisco, CA 94080 (US). (74) Agents: BREZNER, David, J. et al.; Flehr, Hohbach, Test, Albritton &amp; Herbert, Suite 3400, 4 Embarcadero Center, San Francisco, CA 94111-4187 (US).</p>		<p>(81) Designated States: AU, CA, JP, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).  <b>Published</b> <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>
<p>(54) Title: RIP: NOVEL HUMAN PROTEIN INVOLVED IN TUMOR NECROSIS FACTOR SIGNAL TRANSDUCTION, AND SCREENING ASSAYS (57) Abstract The invention relates to a human Receptor Interacting Protein (hRIP), nucleic acids which encode hRIP and methods of using the subject compositions; in particular, methods such as hRIP-based in vitro binding assays and phosphorylation assays for screening chemical libraries for lead compounds for pharmacological agents.</p>		

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*RIP: Novel Human Protein Involved in Tumor Necrosis Factor  
Signal Transduction, and Screening Assays*

## INTRODUCTION

### Field of the Invention

The field of this invention is a novel human kinase involved in tumor necrosis factor signal transduction and its use in drug screening.

### Background

Tumor necrosis factor (TNF) is an important cytokine involved in the signaling of a number of cellular responses including cytotoxicity, anti-viral activity, immun.-regulatory activities and the transcriptional regulation of a number of genes. The TNF receptors (TNF-R1 and TNF-R2) are members of the larger TNF receptor superfamily which also includes the Fas antigen, CD27, CD30, CD40, and the low affinity nerve growth factor receptor. Members of this family have been shown to participate in a variety of biological properties, including programmed cell death, antiviral activity and activation of the transcription factor NF- $\kappa$ B in a wide variety of cell types.

Accordingly, it is desired to identify agents which specifically modulate transduction of TNF receptor family signalling. Unfortunately, the components of the signalling pathway remain largely unknown; hence, the reagents necessary for the development of high-throughput screening assays for such therapeutics are unavailable. Elucidation of TNF receptor family signal transduction pathways leading to NF- $\kappa$ B activation would provide valuable insight into mechanisms to alleviate inflammation. In particular, components of this pathway would provide valuable targets for automated, cost-effective, high throughput drug screening and hence would have immediate application in a broad range of domestic and international pharmaceutical and biotechnology drug development programs.

### Relevant Literature

Stanger et al. (1995) Cell 81, 513-523 report the existence of a Receptor Interacting Protein (RIP) and its functional expression. VanArsdale and Ware (1994) J Immunology 153:3043-3050 describe proteins associated with TNF-R1. The cloning and amino acid sequencing of TNF-R1 is disclosed in Schall et al (1990) Cell 61, 361 and Loetscher et al (1990) Cell 61, 351; the identification of a "death domain" in TNF-R1 is disclosed in Tartaglia et al. (1993) Cell 74:845-853. The cloning and amino acid sequence of a TNF-R associated death domain protein (TRADD) is described by Hsu et al. (1995) Cell 81, 495-504. The cloning and amino acid sequence of the Fas antigen is disclosed in Itoh et al (1991)

Cell 66, 233-243. For a recent review, see Smith et al. (1994) Cell 76:959-962 and Vandenabelle et al. (1995) Trends Cell Biol. 5, 392-399.

### SUMMARY OF THE INVENTION

5           The invention provides methods and compositions relating to a human Receptor Interacting Protein (hRIP). The compositions include nucleic acids which encode hRIP, hRIP kinase domains, and recombinant proteins made from these nucleic acids. The invention also provides methods for screening chemical libraries for lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease associated hRIP activity  
10          or hRIP-dependent signal transduction. In one embodiment, the methods involve incubating a mixture of hRIP, a natural intracellular hRIP substrate or binding target and a candidate pharmacological agent and determining if the presence of the agent modulates the ability of hRIP to selectively phosphorylate the substrate or bind the binding target. Specific agents provide lead compounds for pharmacological agents capable of disrupting hRIP function.

### DETAILED DESCRIPTION OF THE INVENTION

15           A human RIP-encoding nucleic acid sequence is set out in SEQ ID NO: 1. A human RIP kinase domain-encoding nucleic acid sequence is set out in SEQ ID NO: 1, nucleotides 1-900. A human RIP amino acid sequence is set out in SEQ ID NO: 2; and a hRIP kinase  
20          domain sequence is set out in SEQ ID NO:2, residues 1-300.

          Natural nucleic acids encoding hRIP are readily isolated from cDNA libraries with PCR primers and hybridization probes containing portions of the nucleic acid sequence of SEQ ID NO:1. For example, we used low stringency hybridization at 42°C (hybridization  
25          buffer: 20% formamide, 10 % Denhardt, 0.5% SDS, 5X SSPE; with membrane washes at room temperature with 5X SSPE/0.5% SDS) with a 120 base oligonucleotide probe (SEQ ID NO: 1, nucleotides 1728-1847) to isolate a native human RIP cDNA from a library prepared from human umbilical vein endothelial cells. In addition, synthetic hRIP-encoding nucleic acids may be generated by automated synthesis.

          The subject nucleic acids are recombinant, meaning they comprise a sequence joined  
30          to a nucleotide other than that to which sequence is naturally joined and isolated from a natural environment. The nucleic acids may be part of hRIP-expression vectors and may be incorporated into cells for expression and screening, transgenic animals for functional studies

(e.g. the efficacy of candidate drugs for disease associated with expression of a hRIP), etc. These nucleic acids find a wide variety of applications including use as templates for transcription, hybridization probes, PCR primers, therapeutic nucleic acids, etc.; use in detecting the presence of hRIP genes and gene transcripts, in detecting or amplifying nucleic acids encoding additional hRIP homologs and structural analogs, and in gene therapy applications.

The invention provides efficient methods of identifying pharmacological agents or lead compounds for agents active at the level of a hRIP modulatable cellular function, particularly hRIP mediated TNF receptor or Tumor necrosis factor-receptor associated Factor -2 (TRAF2) or TRADD-induced signal transduction. For example, we have found that a binding complex comprising TNF R1, TRADD, and hRIP exists in TNF-stimulated cells. Generally, the screening methods involve assaying for compounds which interfere with a hRIP activity such as kinase activity or TRAF2 or TRADD binding. The methods are amenable to automated, cost-effective high throughput screening of chemical libraries for lead compounds. Identified reagents find use in the pharmaceutical industries for animal and human trials; for example, the reagents may be derivatized and rescreened in in vitro and in vivo assays to optimize activity and minimize toxicity for pharmaceutical development. Target therapeutic indications are limited only in that the target cellular function be subject to modulation, usually inhibition, by disruption of the formation of a complex comprising hRIP and one or more natural hRIP intracellular binding targets including substrates or otherwise modulating hRIP kinase activity. Target indications may include infection, genetic disease, cell growth and regulatory or immunologic dysfunction, such as neoplasia, inflammation, hypersensitivity, etc.

A wide variety of assays for binding agents are provided including labeled in vitro kinase assays, protein-protein binding assays, immunoassays, cell based assays, etc. The hRIP compositions used in the methods are recombinantly produced from nucleic acids having the disclosed hRIP nucleotide sequences. The hRIP may be part of a fusion product with another peptide or polypeptide, e.g. a polypeptide that is capable of providing or enhancing protein-protein binding, stability under assay conditions (e.g. a tag for detection or anchoring), etc.

The assay mixtures comprise one or more natural intracellular hRIP binding targets including substrates, such as TRADD, TRAF2, or, in the case of an autophosphorylation

assay, the hRIP itself can function as the binding target. In one embodiment, the mixture comprises a complex of hRIP, TRADD and TNFR1. A hRIP derived pseudosubstrate may be used or modified (e.g. A to S/T substitutions) to generate effective substrates for use in the subject kinase assays as can synthetic peptides or other protein substrates. Generally, hRIP-specificity of the binding agent is shown by kinase activity (i.e. the agent demonstrates activity of an hRIP substrate, agonist, antagonist, etc.) or binding equilibrium constants (usually at least about  $10^6 \text{ M}^{-1}$ , preferably at least about  $10^8 \text{ M}^{-1}$ , more preferably at least about  $10^9 \text{ M}^{-1}$ ). A wide variety of cell-based and cell-free assays may be used to demonstrate hRIP-specific binding; preferred are rapid in vitro, cell-free assays such as mediating or inhibiting hRIP-protein (e.g. hRIP-TRADD) binding, phosphorylation assays, immunoassays, etc.

The assay mixture also comprises a candidate pharmacological agent. Candidate agents encompass numerous chemical classes, though typically they are organic compounds; preferably small organic compounds and are obtained from a wide variety of sources including libraries of synthetic or natural compounds. A variety of other reagents may also be included in the mixture. These include reagents like salts, buffers, neutral proteins, e.g. albumin, detergents, etc. which may be used to facilitate optimal binding and/or reduce non-specific or background interactions, etc. Also, reagents that otherwise improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, antimicrobial agents, etc. may be used.

In a preferred in vitro, binding assay, a mixture of at least the kinase domain of hRIP, one or more binding targets or substrates and the candidate agent is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the hRIP specifically binds the cellular binding target at a first binding affinity or phosphorylates the substrate at a first rate. After incubation, a second binding affinity or rate is detected. Detection may be effected in any convenient way. For cell-free binding assays, one of the components usually comprises or is coupled to a label. The label may provide for direct detection as radioactivity, luminescence, optical or electron density, etc. or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods may be used to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and thereafter the label detected.

The following examples are offered by way of illustration and not by way of limitation.

### EXAMPLES

5 1. Protocol for hRIP autophosphorylation assay.

A. Reagents:

- Neutralite Avidin: 20 µg/ml in PBS.

- hRIP:  $10^{-8}$  -  $10^{-5}$  M biotinylated hRIP kinase domain, residues 1-300 at 20 µg/ml in PBS.

10 - Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.

- [ $^{32}$ P]γ-ATP 10x stock:  $2 \times 10^{-5}$  M cold ATP with 100 µCi [ $^{32}$ P]γ-ATP. Place in the 4°C microfridge during screening.

15 - Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO<sub>3</sub> (Sigma # S-6508) in 10 ml PBS.

B. Preparation of assay plates:

20 - Coat with 120 µl of stock Neutralite avidin per well overnight at 4°C.

- Wash 2 times with 200 µl PBS.

- Block with 150 µl of blocking buffer.

- Wash 2 times with 200 µl PBS.

C. Assay:

25 - Add 40 µl assay buffer/well.

- Add 40 µl biotinylated hRIP (0.1-10 pmoles/40 ul in assay buffer)

- Add 10 µl compound or extract.

- Add 10 µl [ $^{32}$ P]γ-ATP 10x stock.

- Shake at 30°C for 15 minutes.

30 - Incubate additional 45 minutes at 30°C.

- Stop the reaction by washing 4 times with 200 µl PBS.

- Add 150 µl scintillation cocktail.

- Count in Topcount.

D. Controls for all assays (located on each plate):

- a. Non-specific binding (no RIP added)
- b. cold ATP to achieve 80% inhibition.

5

2. Protocol for hRIP - substrate phosphorylation assay.

A. Reagents:

- Neutralite Avidin: 20 µg/ml in PBS.

- hRIP:  $10^{-8}$  -  $10^{-5}$  M hRIP at 20 µg/ml in PBS.

- 10
- Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.

- [ $^{32}$ P]γ-ATP 10x stock:  $2 \times 10^{-5}$  M cold ATP with 100 µCi [ $^{32}$ P]γ-ATP. Place in the 4°C microfridge during screening.

- 15
- Substrate:  $2 \times 10^{-6}$  M biotinylated synthetic peptide kinase substrate at 20 µg/ml in PBS.

- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO<sub>3</sub> (Sigma # S-6508) in 10 ml PBS.

20

B. Preparation of assay plates:

- Coat with 120 µl of stock Neutralite avidin per well overnight at 4°C.

- Wash 2 times with 200 µl PBS.

- Block with 150 µl of blocking buffer.

- 25
- Wash 2 times with 200 µl PBS.

C. Assay:

- Add 40 µl assay buffer/well.

- Add 40 µl hRIP (0.1-10 pmoles/40 ul in assay buffer)

- Add 10 µl compound or extract.

- 30
- Shake at 30°C for 15 minutes.

- Add 10 µl [ $^{32}$ P]γ-ATP 10x stock.

- Add 10 µl substrate.



- Shake at 30°C for 15 minutes.
- Incubate additional 45 minutes at 30°C.
- Stop the reaction by washing 4 times with 200 µl PBS.
- Add 150 µl scintillation cocktail.
- Count in Topcount.

- 5
- D. Controls for all assays (located on each plate):
- a. Non-specific binding (no RIP added)
  - b. cold ATP to achieve 80% inhibition.

10 3. Protocol for hRIP - TRADD binding assay.

A. Reagents:

- Anti-myc antibody: 20 µg/ml in PBS.
- Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol,
- 15 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.

- <sup>32</sup>P hRIP 10x stock:  $10^{-8}$  -  $10^{-6}$  M "cold" hRIP (full length) supplemented with 200,000-250,000 cpm of labeled hRIP (HMK-tagged) (Beckman counter). Place in the 4°C microfridge during screening.

- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO<sub>3</sub> (Sigma # S-6508) in 10 ml PBS.

- TRADD:  $10^{-8}$  -  $10^{-5}$  M myc epitope-tagged TRADD in PBS.

B. Preparation of assay plates:

- 25
- Coat with 120 µl of stock anti-myc antibody per well overnight at 4°C.
  - Wash 2X with 200 µl PBS.
  - Block with 150 µl of blocking buffer.
  - Wash 2X with 200 µl PBS.

C. Assay:

- 30
- Add 40 µl assay buffer/well.
  - Add 10 µl compound or extract.
  - Add 10 µl <sup>33</sup>P-RIP (20,000-25,000 cpm/0.1-10 pmoles/well =  $10^{-9}$  -  $10^{-7}$  M final

concentration).

- Shake at 25°C for 15 minutes.
- Incubate additional 45 minutes at 25°C.
- Add 40 µl epitope-tagged TRADD (0.1-10 pmoles/40 ul in assay buffer)
- Incubate 1 hour at room temperature.
- Stop the reaction by washing 4 times with 200 µl PBS.
- Add 150 µl scintillation cocktail.
- Count in Topcount.

D. Controls for all assays (located on each plate):

- a. Non-specific binding (no hRIP added)
- b. Soluble (non-tagged TRADD) to achieve 80% inhibition.

4. Protocol for hRIP - TRAF2 binding assay.

A. Reagents:

- Anti-myc antibody: 20 µg/ml in PBS.
- Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.
- Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.

- <sup>32</sup>P hRIP 10x stock:  $10^{-8}$  -  $10^{-6}$  M "cold" hRIP kinase domain, residues 1-300, supplemented with 200,000-250,000 cpm of labeled hRIP kinase domain (HMK-tagged) (Beckman counter). Place in the 4°C microfridge during screening.

- Protease inhibitor cocktail (1000X): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB # 236624), 25 mg Benzamidine (Sigma # B-6506), 25 mg Leupeptin (BMB # 1017128), 10 mg APMSF (BMB # 917575), and 2mM NaVO<sub>3</sub> (Sigma # S-6508) in 10 ml PBS.

- TRAF2:  $10^{-8}$  -  $10^{-5}$  M myc epitope-tagged TRAF2 in PBS.

B. Preparation of assay plates:

- Coat with 120 µl of stock anti-myc antibody per well overnight at 4°C.
- Wash 2X with 200 µl PBS.
- Block with 150 µl of blocking buffer.
- Wash 2X with 200 µl PBS.

C. Assay:

- Add 40  $\mu$ l assay buffer/well.
- Add 10  $\mu$ l compound or extract.
- Add 10  $\mu$ l  $^{33}$ P-RIP kinase domain (20,000-25,000 cpm/0.1-10 pmoles/well =  $10^{-9}$ -  $10^{-7}$  M final concentration).

5

- Shake at 25°C for 15 minutes.
- Incubate additional 45 minutes at 25°C.
- Add 40  $\mu$ l epitope-tagged TRAF2 (0.1-10 pmoles/40  $\mu$ l in assay buffer)
- Incubate 1 hour at room temperature.
- Stop the reaction by washing 4 times with 200  $\mu$ l PBS.

10

- Add 150  $\mu$ l scintillation cocktail.
- Count in Topcount.

D. Controls for all assays (located on each plate):

- a. Non-specific binding (no hRIP kinase domain added)
- b. Soluble (non-tagged TRAF2) to achieve 80% inhibition.

15

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

20

## SEQUENCE LISTING

## (1) GENERAL INFORMATION:

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(ii) TITLE OF INVENTION: RIP: NOVEL HUMAN PROTEIN INVOLVED IN  
TUMOR NECROSIS FACTOR SIGNAL TRANSDUCTION, AND SCREENING  
ASSAYS

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(F) ZIP: 94111-4187

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk

(B) COMPUTER: IBM PC compatible

(C) OPERATING SYSTEM: PC-DOS/MS-DOS

(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:

(B) FILING DATE:

(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:

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(B) REGISTRATION NUMBER: 24,774

(C) REFERENCE/DOCKET NUMBER: T95-006/PCT

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## (2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 2016 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..2013

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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5	TTC CTG GAG AGT GCA GAA CTG GAC AGC GGA GGC TTT GGG AAG GTG TCT	96
	Phe Leu Glu Ser Ala Glu Leu Asp Ser Gly Gly Phe Gly Lys Val Ser	
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	Leu Cys Phe His Arg Thr Gln Gly Leu Met Ile Met Lys Thr Val Tyr	
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	275 280 285	
10	TAT TTA AGT CAA TTA GAA GAA AGT GTA GAA GAG GAC GTG AAG AGT TTA	912
	Tyr Leu Ser Gln Leu Glu Glu Ser Val Glu Glu Asp Val Lys Ser Leu	
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	355 360 365	
25	GAG AAT GAG CCC AGC CTG CAG AGT AAA CTC CAA GAC GAA GCC AAC TAC	1152
	Glu Asn Glu Pro Ser Leu Gln Ser Lys Leu Gln Asp Glu Ala Asn Tyr	
	370 375 380	
	CAT CTT TAT GGC AGC CGC ATG GAC AGG CAG ACG AAA CAG CAG CCC AGA	1200
	His Leu Tyr Gly Ser Arg Met Asp Arg Gln Thr Lys Gln Gln Pro Arg	
30	385 390 395 400	
	CAG AAT GTG GCT TAC AAC AGA GAG GAG GAA AGG AGA CGC AGG GTC TCC	1248
	Gln Asn Val Ala Tyr Asn Arg Glu Glu Glu Arg Arg Arg Arg Val Ser	
	405 410 415	
	CAT GAC CCT TTT GCA CAG CAA AGA CCT TAC GAG AAT TTT CAG AAT ACA	1296
35	His Asp Pro Phe Ala Gln Gln Arg Pro Tyr Glu Asn Phe Gln Asn Thr	
	420 425 430	
	GAG GGA AAA GGC ACT GTT TAT TCC AGT GCA GCC AGT CAT GGT AAT GCA	1344
	Glu Gly Lys Gly Thr Val Tyr Ser Ser Ala Ala Ser His Gly Asn Ala	
	435 440 445	
40	GTG CAC CAG CCC TCA GGG CTC ACC AGC CAA CCT CAA GTA CTG TAT CAG	1392
	Val His Gln Pro Ser Gly Leu Thr Ser Gln Pro Gln Val Leu Tyr Gln	
	450 455 460	
	AAC AAT GGA TTA TAT AGC TCA CAT GGC TTT GGA ACA AGA CCA CTG GAT	1440
	Asn Asn Gly Leu Tyr Ser Ser His Gly Phe Gly Thr Arg Pro Leu Asp	
45	465 470 475 480	

	CCA GGA ACA GCA GGT CCC AGA GTT TGG TAC AGG CCA ATT CCA AGT CAT	1488
	Pro Gly Thr Ala Gly Pro Arg Val Trp Tyr Arg Pro Ile Pro Ser His	
	485 490 495	
5	ATG CCT AGT CTG CAT AAT ATC CCA GTG CCT GAG ACC AAC TAT CTA GGA	1536
	Met Pro Ser Leu His Asn Ile Pro Val Pro Glu Thr Asn Tyr Leu Gly	
	500 505 510	
	AAT ACA CCC ACC ATG CCA TTC AGC TCC TTG CCA CCA ACA GAT GAA TCT	1584
	Asn Thr Pro Thr Met Pro Phe Ser Ser Leu Pro Pro Thr Asp Glu Ser	
	515 520 525	
10	ATA AAA TAT ACC ATA TAC AAT AGT ACT GGC ATT CAG ATT GGA GCC TAC	1632
	Ile Lys Tyr Thr Ile Tyr Asn Ser Thr Gly Ile Gln Ile Gly Ala Tyr	
	530 535 540	
	AAT TAT ATG GAG ATT GGT GGG ACG AGT TCA TCA CTA CTA GAC AGC ACA	1680
	Asn Tyr Met Glu Ile Gly Gly Thr Ser Ser Ser Leu Leu Asp Ser Thr	
15	545 550 555 560	
	AAT ACG AAC TTC AAA GAA GAG CCA GCT GCT AAG TAC CAA GCT ATC TTT	1728
	Asn Thr Asn Phe Lys Glu Glu Pro Ala Ala Lys Tyr Gln Ala Ile Phe	
	565 570 575	
	GAT AAT ACC ACT AGT CTG ACG GAT AAA CAC CTG GAC CCA ATC AGG GAA	1776
20	Asp Asn Thr Thr Ser Leu Thr Asp Lys His Leu Asp Pro Ile Arg Glu	
	580 585 590	
	AAT CTG GGA AAG CAC TGG AAA AAC TGT GCC CGT AAA CTG GGC TTC ACA	1824
	Asn Leu Gly Lys His Trp Lys Asn Cys Ala Arg Lys Leu Gly Phe Thr	
	595 600 605	
25	CAG TCT CAG ATT GAT GAA ATT GAC CAT GAC TAT GAG CGA GAT GGA CTG	1872
	Gln Ser Gln Ile Asp Glu Ile Asp His Asp Tyr Glu Arg Asp Gly Leu	
	610 615 620	
	AAA GAA AAG GTT TAC CAG ATG CTC CAA AAG TGG GTG ATG AGG GAA GGC	1920
	Lys Glu Lys Val Tyr Gln Met Leu Gln Lys Trp Val Met Arg Glu Gly	
30	625 630 635 640	
	ATA AAG GGA GCC ACG GTG GGG AAG CTG GCC CAG GCG CTC CAC CAG TGT	1968
	Ile Lys Gly Ala Thr Val Gly Lys Leu Ala Gln Ala Leu His Gln Cys	
	645 650 655	
	TCC AGG ATC GAC CTT CTG AGC AGC TTG ATT TAC GTC AGC CAG AAC	2013
35	Ser Arg Ile Asp Leu Leu Ser Ser Leu Ile Tyr Val Ser Gln Asn	
	660 665 670	
	TAA	2016

## (2) INFORMATION FOR SEQ ID NO:2:

40

## (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 671 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

	Met	Gln	Pro	Asp	Met	Ser	Leu	Asn	Val	Ile	Lys	Met	Lys	Ser	Ser	Asp
	1				5					10					15	
	Phe	Leu	Glu	Ser	Ala	Glu	Leu	Asp	Ser	Gly	Gly	Phe	Gly	Lys	Val	Ser
				20					25					30		
5	Leu	Cys	Phe	His	Arg	Thr	Gln	Gly	Leu	Met	Ile	Met	Lys	Thr	Val	Tyr
		35						40					45			
	Lys	Gly	Pro	Asn	Cys	Ile	Glu	His	Asn	Glu	Ala	Leu	Leu	Glu	Glu	Ala
		50					55					60				
10	Lys	Met	Met	Asn	Arg	Leu	Arg	His	Ser	Arg	Val	Val	Lys	Leu	Leu	Gly
	65				70					75						80
	Val	Ile	Ile	Glu	Glu	Gly	Lys	Tyr	Ser	Leu	Val	Met	Glu	Tyr	Met	Glu
				85						90					95	
	Lys	Gly	Asn	Leu	Met	His	Val	Leu	Lys	Ala	Glu	Met	Ser	Thr	Pro	Leu
				100					105					110		
15	Ser	Val	Lys	Gly	Arg	Ile	Ile	Trp	Glu	Ile	Ile	Glu	Gly	Met	Cys	Tyr
		115						120					125			
	Leu	His	Gly	Lys	Gly	Val	Ile	His	Lys	Asp	Leu	Lys	Pro	Glu	Asn	Ile
		130					135					140				
20	Leu	Val	Asp	Asn	Asp	Phe	His	Ile	Lys	Ile	Ala	Asp	Leu	Gly	Leu	Ala
	145					150					155					160
	Ser	Phe	Lys	Met	Trp	Ser	Lys	Leu	Asn	Asn	Glu	Glu	His	Asn	Glu	Leu
				165					170					175		
	Arg	Glu	Val	Asp	Gly	Thr	Ala	Lys	Lys	Asn	Gly	Gly	Thr	Leu	Tyr	Tyr
				180					185					190		
25	Met	Ala	Pro	Glu	His	Leu	Asn	Asp	Val	Asn	Ala	Lys	Pro	Thr	Glu	Lys
		195						200					205			
	Ser	Asp	Val	Tyr	Ser	Phe	Ala	Val	Val	Leu	Trp	Ala	Ile	Phe	Ala	Asn
		210				215					220					
30	Lys	Glu	Pro	Tyr	Glu	Asn	Ala	Ile	Cys	Glu	Gln	Gln	Leu	Ile	Met	Cys
	225					230				235					240	
	Ile	Lys	Ser	Gly	Asn	Arg	Pro	Asp	Val	Asp	Asp	Ile	Thr	Glu	Tyr	Cys
				245					250					255		
	Pro	Arg	Glu	Ile	Ile	Ser	Leu	Met	Lys	Leu	Cys	Trp	Glu	Ala	Asn	Pro
				260					265					270		
35	Glu	Ala	Arg	Pro	Thr	Phe	Pro	Gly	Ile	Glu	Glu	Lys	Phe	Arg	Pro	Phe
		275						280					285			
	Tyr	Leu	Ser	Gln	Leu	Glu	Glu	Ser	Val	Glu	Glu	Asp	Val	Lys	Ser	Leu
		290				295						300				
40	Lys	Lys	Glu	Tyr	Ser	Asn	Glu	Asn	Ala	Val	Val	Lys	Arg	Met	Gln	Ser
	305					310				315					320	
	Leu	Gln	Leu	Asp	Cys	Val	Ala	Val	Pro	Ser	Ser	Arg	Ser	Asn	Ser	Ala
				325					330					335		
	Thr	Glu	Gln	Pro	Gly	Ser	Leu	His	Ser	Ser	Gln	Gly	Leu	Gly	Met	Gly
45				340					345					350		



	Pro	Val	Glu	Glu	Ser	Trp	Phe	Ala	Pro	Ser	Leu	Glu	His	Pro	Gln	Glu	
			355					360					365				
	Glu	Asn	Glu	Pro	Ser	Leu	Gln	Ser	Lys	Leu	Gln	Asp	Glu	Ala	Asn	Tyr	
		370					375					380					
5	His	Leu	Tyr	Gly	Ser	Arg	Met	Asp	Arg	Gln	Thr	Lys	Gln	Gln	Pro	Arg	
	385					390					395					400	
	Gln	Asn	Val	Ala	Tyr	Asn	Arg	Glu	Glu	Glu	Arg	Arg	Arg	Arg	Val	Ser	
					405					410					415		
	His	Asp	Pro	Phe	Ala	Gln	Gln	Arg	Pro	Tyr	Glu	Asn	Phe	Gln	Asn	Thr	
10				420					425					430			
	Glu	Gly	Lys	Gly	Thr	Val	Tyr	Ser	Ser	Ala	Ala	Ser	His	Gly	Asn	Ala	
			435					440					445				
	Val	His	Gln	Pro	Ser	Gly	Leu	Thr	Ser	Gln	Pro	Gln	Val	Leu	Tyr	Gln	
		450					455					460					
15	Asn	Asn	Gly	Leu	Tyr	Ser	Ser	His	Gly	Phe	Gly	Thr	Arg	Pro	Leu	Asp	
	465					470					475					480	
	Pro	Gly	Thr	Ala	Gly	Pro	Arg	Val	Trp	Tyr	Arg	Pro	Ile	Pro	Ser	His	
					485					490					495		
	Met	Pro	Ser	Leu	His	Asn	Ile	Pro	Val	Pro	Glu	Thr	Asn	Tyr	Leu	Gly	
20				500					505					510			
	Asn	Thr	Pro	Thr	Met	Pro	Phe	Ser	Ser	Leu	Pro	Pro	Thr	Asp	Glu	Ser	
				515					520					525			
	Ile	Lys	Tyr	Thr	Ile	Tyr	Asn	Ser	Thr	Gly	Ile	Gln	Ile	Gly	Ala	Tyr	
		530					535					540					
25	Asn	Tyr	Met	Glu	Ile	Gly	Gly	Thr	Ser	Ser	Ser	Leu	Leu	Asp	Ser	Thr	
	545					550					555					560	
	Asn	Thr	Asn	Phe	Lys	Glu	Glu	Pro	Ala	Ala	Lys	Tyr	Gln	Ala	Ile	Phe	
					565					570					575		
	Asp	Asn	Thr	Thr	Ser	Leu	Thr	Asp	Lys	His	Leu	Asp	Pro	Ile	Arg	Glu	
30				580					585					590			
	Asn	Leu	Gly	Lys	His	Trp	Lys	Asn	Cys	Ala	Arg	Lys	Leu	Gly	Phe	Thr	
			595					600					605				
	Gln	Ser	Gln	Ile	Asp	Glu	Ile	Asp	His	Asp	Tyr	Glu	Arg	Asp	Gly	Leu	
		610					615					620					
35	Lys	Glu	Lys	Val	Tyr	Gln	Met	Leu	Gln	Lys	Trp	Val	Met	Arg	Glu	Gly	
	625					630					635					640	
	Ile	Lys	Gly	Ala	Thr	Val	Gly	Lys	Leu	Ala	Gln	Ala	Leu	His	Gln	Cys	
					645					650					655		
	Ser	Arg	Ile	Asp	Leu	Leu	Ser	Ser	Leu	Ile	Tyr	Val	Ser	Gln	Asn		
40				660					665					670			

WHAT IS CLAIMED IS:

1. An isolated, recombinant nucleic acid encoding a human Receptor Interacting Protein (hRIP) kinase domain.

2. An isolated, recombinant nucleic acid encoding a human Receptor Interacting Protein (hRIP) comprising SEQ ID NO: 1.

3. A method of making a human Receptor Interacting Protein (hRIP) kinase domain containing protein, said method comprising the steps of translating a nucleic acid according to claim 1 to form a translation product and isolating said translation product.

4. A method of identifying lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease, said method comprising the steps of:

making a protein according to the method of claim 3,

forming a mixture comprising:

said protein,

a natural intracellular hRIP binding target, wherein said binding target is capable of specifically binding said protein, and

a candidate pharmacological agent;

incubating said mixture under conditions whereby, but for the presence of said candidate pharmacological agent, said protein selectively binds said binding target at a first binding affinity;

detecting a second binding affinity of said protein to said binding target,

wherein a difference between said first and second binding affinity indicates that said candidate pharmacological agent is a lead compound for a pharmacological agent capable of modulating hRIP-dependent signal transduction.

5. A method according to claim 4, wherein said hRIP binding target comprises a Tumor necrosis factor receptor Associated Factor -2 (TRAF2) or a Tumor necrosis factor Receptor-1 Associated Death Domain protein (TRADD).

6. A method of identifying lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease, said method comprising the steps of:

making a protein according to the method of claim 3,

forming a mixture comprising:

5           said protein,

an hRIP substrate, wherein said hRIP kinase domain of said protein is capable of specifically phosphorylating said substrate, and

a candidate pharmacological agent;

10           incubating said mixture under conditions whereby, but for the presence of said candidate pharmacological agent, said hRIP kinase domain selectively phosphorylates said substrate at a first rate;

detecting a second rate of phosphorylation of said substrate by said hRIP kinase domain,

15           wherein a difference between said first and second rate indicates that said candidate pharmacological agent is a lead compound for a pharmacological agent capable of modulating hRIP kinase activity.

7. A method according to claim 6 wherein said hRIP substrate is hRIP.

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US96/16778

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.  
US CL : 536/23.5; 435/ 69.1, 69.5, 252.3, 320.1; 530/350, 351  
According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/23.5; 435/ 69.1, 69.5, 252.3, 320.1; 530/350, 351

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ---	STANGER et al. RIP: A Novel Protein Containing a Death Domain That Interacts with Fas/APO-1 (CD95) in Yeast and Causes Cell Death. Cell. 19 May 1995, Vol. 81, pages 513-523, see Figs. 2-3, and sequence alignment,	1-3 -----
Y		2
Y, P	WO 96/25941 A1 (YEDA RESEARCH & DEVELOPMENT CO. LTD.) 29 August 1996 (29/08/96), see abstract, figures and claims.	1-3
A	HSU et al. The TNF Receptor 1-Associated Protein TRADD Signals Cell Death and NF-kB Activation. Cell, 19 May 1995, Vol. 81, pages 495-504, see all.	1-3

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	* T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* A		document defining the general state of the art which is not considered to be of particular relevance
* E		earlier document published on or after the international filing date
* L		document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reasons (as specified)
* O		document referring to an oral disclosure, use, exhibition or other means
* P		document published prior to the international filing date but later than the priority date claimed
	* X	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
	* Y	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
	* &	document member of the same patent family

Date of the actual completion of the international search  
15 JANUARY 1997

Date of mailing of the international search report  
28 FEB 1997

Name and mailing address of the ISA/US  
Commissioner of Patents and Trademarks  
Box PCT  
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Facsimile No. (703) 305-3230

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## INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US96/16778

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A, P	BAKER et al. Transducers of Life and Death: TNF Receptor Superfamily and Associated Proteins. Oncogene, 04 January 1996, Vol. 12, pages 1-9, see all	1-3

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US96/16778

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2. ☐ Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3. ☐ Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  
1-3

Remark on Protest

☐  
☐

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

**A. CLASSIFICATION OF SUBJECT MATTER:**  
IPC (6):

C07H 21/04; C12P 21/06, 21/02; C12N 1/20, 15/00; C07K 1/00, 14/52

**BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING**  
This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claims 1-3, drawn to nucleic acids that encode for human Receptor Interacting Proteins (hRIP) and methods of making the encoded proteins.

Group II, claims 4-7, drawn to methods of identifying lead compounds.

The inventions listed as Groups do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: The special technical feature of Group I is directed to nucleic acids that encode for hRIP and to methods of making hRIP; whereas the special technical feature of Group II is directed to methods of identifying lead compounds. The methods of these two groups do not share a special technical and unifying feature, because each of these methods require the utilization of different process/method steps, different elements/agents, and their are different starting material and the final outcomes are also different. Furthermore, these methods and their steps and elements are not required one for the other.